

Enzyme Production and Biotypes of Vaginal *Candida albicans*

ZEFIRYN CYBULSKI¹, ELŻBIETA KRZEMIŃSKA-JAŚKOWIAK, PRZEMYSŁAW MAJEWSKI²,
JERZY CHYLAK³ and MAREK PAWLIK⁴

¹ Wielkopolska Cancer Center, ² Department of Clinical Pathomorphology,
³ Department of Medical Microbiology, University of Medical Sciences, Poznań,
⁴ Central Hospital, Lutycka str., Poznań, Poland

Received 28 December 2004, received in revised form 4 March 2005, accepted 17 June 2005

Abstract

Candidal vulvovaginitis is one of the most common forms of vaginal infection. However, the origin of the infecting organism is sometimes doubtful. Therefore, epidemiological investigation can help to recognize routes of infection spreading. The aim of the present study was to determine the ability to produce esterases by clinical isolates of *C. albicans* and to find the relationship between their serotypes. Also, it was intended to determine the ability of these strains to produce proteases and lipases as well as the ability of the strains to assimilate carbohydrates. 46 strains of *C. albicans* isolates from the vagina of women suffering from vulvovaginitis were examined. Three main kinds of esterases were distinguished by their spectra of hydrolytic activity toward α -naphthyl acetate, β -naphthyl propionate and indoxyl acetate. The strains were grouped into four categories: three categories in which esterase patterns were observed and one category in which esterase bands were not observed. On the basis of the 20 carbon sources assimilated, the *C. albicans* strains were categorized into 11 biotypes with the major biotype accounting for 21 (45.7%) strains. The examination of proteolytic activity using casein and albumin enabled to divide the strains into four groups. All of the examined strains belonged to serotype A and all of them expressed lipolytic activity. Esterase electrophoretic patterns and biotypes based on proteolytic activities were compared with the ability to assimilate carbon from various sources.

Key words: *C. albicans*, vulvovaginitis, biotypes, esterases

Introduction

The use of epidemiological markers to type *C. albicans* in communicable diseases is recognized as being very important for the following purposes: detection of sources and routes of infections; distinction between recurrent and chronic infection; identification of endemic strains; selection of specific immunoprophylaxis and immunotherapy (al Rawi and Kavanagh, 1999; Kubota, 1998; Magliani *et al.*, 2002; Mendling *et al.*, 2000; Moraes *et al.*, 2000). In the case of *Candida* vulvovaginitis, the origin of the infecting organism is unclear. It has been suggested that there are many women who carry *C. albicans* in the vagina without subjective symptomatology, often with low *Candida* concentration. These observations are in agreement with the view that *C. albicans* may be either human commensal or a pathogen (Kubota, 1998; Marai, 2001). *C. albicans* is a common opportunistic pathogen in HIV-infected patients. Diabetes mellitus increases the rate of vaginal colonization and infection with *C. albicans* spp. (Goswami *et al.*, 2000; Haberland-Carrodeguas *et al.*, 2002; Taylor *et al.*, 2000). However, *C. albicans* is a potent allergen in some situations and it has been suggested that local hypersensitivity to these fungi is a factor which prolongs recurrent vaginal candidiasis (Moraes *et al.*, 2000).

Various methods have been used to type *Candida* isolates, with the objective of developing epidemiological tools. The most widely used methods include biotyping, serological typing, comparison of the susceptibility or resistance response to antifungal chemicals, and the ability of the strains to produce hydrolytic enzymes (Kantarcioglu 2002; Kurnatowska, 1998; Mendling *et al.*, 2000; Mercure *et al.*, 1996; Quindos *et al.*, 1996).

Hydrolytic enzymes of *C. albicans* have been implicated as virulence factors, and particular emphasis has been given to the extracellular protease, esterase and lipase (De Bernardis *et al.*, 1999; Kantarcioglu 2002; Kurnatowska, 1998). The goals of the present study were: firstly, to determine the patterns of esterase

electrophoretic types (zymotypes) among clinical isolates of vaginal *C. albicans*, secondly, to find out whether the zymotypes could be used to define strains of *C. albicans* for epidemiological purposes, thirdly, to determine the serotype of the strains. Moreover, the aim was to compare *C. albicans* strains with respect to their esterase patterns, their capability of secreting protease and lipase and their ability to assimilate carbohydrates.

Experimental

Materials and Methods

46 strains of *Candida albicans* were isolated from vagina of women suffering from vulvovaginitis. The age of patients ranged from 17 to 67 years. Samples were taken from the wall or fluid (pool) of the vaginal canal with a sterile cotton swab and immediately streaked on Sabouraud glucose plates agar. Isolates were found to be *C. albicans* if they showed germ tube production in rabbit serum and if they were able to produce chlamydo spores on Nickerson-Mankowski medium. Apart from that, strains of *Candida albicans* were typed using API 20C AUX (bioMerieux). Proteolytic activities of the strains examined were determined using human albumin and bovine casein as substrates according to the procedure described by Staib (1965). Lipolytic activities were determined using the medium containing Tween 80 according to Werner (1966). API 20 C AUX test was performed according to the manufacturer's instruction. For each of the isolates the serotype was determined by slide agglutination test according to Hannula *et al.* (2001). The determination of the esterase patterns was performed using the techniques described by Goulet and Picard (1991) and Branger *et al.* (1990) which were adapted in order to examine strains of *C. albicans*. The single colony of each examined strain of *C. albicans* growing on Sabouraud agar was inoculated into Sabouraud broth then used to determine the esterase pattern.

C. albicans was grown overnight at 37°C in 40 ml of Sabouraud broth in Erlenmeyer flasks being vigorously shaken. After centrifugation the yeasts were washed in 0.075 M Tris-0.06 M glycine buffer (pH 8,7), resuspended in 1.5 ml of the same buffer and then disrupted 10 times by refrigeration and thawing. Debris was removed at 10.000×g for 15 min. at 4°C. The supernatants containing at least 6 mg of protein per 1 ml were stored at -20°C until used for electrophoresis. Each strain was cultured and extracted at least twice. Horizontal slab polyacrylamide-agarose gel electrophoresis and an estimation of electrophoretic mobility were performed as described by Goulet and Picard, 1991 and Uriel 1966. The detection of hydrolytic activity to a naphthyl acetate, b naphthyl propionate and indoxyl acetate was carried out by staining with the dye Fast Blue.

Results

All of the examined strains belonged to serotype A and all of them showed lipolytic activity. The examination of esterase electrophoretic motility allowed to differentiate the examined strains into four groups of which one group was constituted by strains not producing esterases detected in the present study. All of the

three substrates used: α -naphthyl acetate, β -naphthyl propionate and indoxyl acetate, were hydrolysed by fungal esterases. 15 of the *C. albicans* strains exhibited an esterase pattern consisting of two distinctly stained bands. 7 strains showed a simple esterase pattern consisting of a single band and the pattern of 5 strains consisted of three bands (zymotypes A, B, C, respectively). The mobility of these enzymes and esterase patterns of the examined strains are shown in Fig. 1. 19 strains of *C. albicans* did not exhibit esterase activity detected in this examination.

Basing on proteolytic activities, the strains were divided into four groups. 23 of 46 examined strains exhibited proteolytic activity to human albumin and bovine casein, 10 strains were active with respect to only bovine casein and 11 strains were active only with respect to human albumin. 2 strains did not exhibit proteolytic activity with respect to both substrates used (Table I).

On the basis of 20 carbon sources assimilated, the *C. albicans* isolates were categorized into 11 biotypes with the major biotype accounting for 21 (45.7%) strains of numerical profile 2 576 174. 13 of them hydrolyzed both substrates: human albumin and bovine casein. 8 of the strains from these 21 strains belonged to pattern A of esterase production. The relationship between esterase

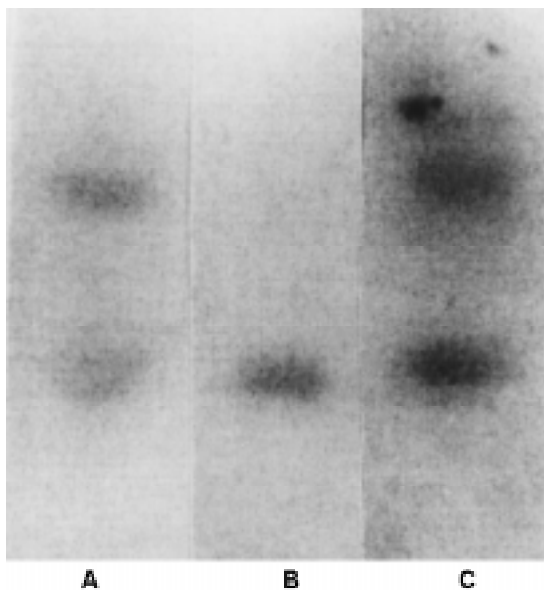


Fig. 1. Esterase patterns of *C. albicans*

Electrophoretic mobility of *Candida albicans* esterases in horizontal slab polyacrylamide – agarose gel electrophoresis. Three esterase patterns of *C. albicans* are presented. All patterns show esterases which hydrolysed α -naphthyl acetate, β -naphthyl propionate and indoxyl acetate. 27 of 46 examined *C. albicans* strains exhibited esterase activity detectable in used method.

Table I
Biotypes according to proteolytic activity of *C. albicans*

Biotype	Proteolytic activity		Number of strains
	H.A. ^a	B.C. ^b	
I	+	+	23
I	–	+	10
III	+	–	11
IV	–	–	2

H.A.^a– human albumin, B.C.^b– bovine casein
(+) – presence of proteolysis
(–) – absence of proteolysis

Table II
The comparison of 21 strains of *C. albicans* exhibiting the same biochemical properties

Numerical profile (2576174) of 21 strains examined	Proteolytic activity with respect to			Lack of proteolysis	Esterase pattern			
	HA ^a	BC ^b	HA&BC		A	B	C	0
No of strains	3	4	13	1	8	2	2	9

HA^a human albumin, BC^b bovine casein
A, B, C – zymogram characterized by two, one and three esterase bands, respectively
0 – lack of esterase pattern.

patterns and proteolytic activity of the strains exhibiting the most common biotype is shown in Table II. The remaining 25 strains belonging to ten different biotypes showed a variety of esterase patterns and they differed in their proteolytic activities to the substrates used, making any comparison impossible.

Discussion

Candida albicans is an important cause of a wide spectrum of diseases including vulvovaginitis (Farina *et al.*, 2000; Linhares *et al.*, 2001; Marai, 2001; Mendling *et al.*, 2000). The presence of *Candida* vulvovaginitis cannot be definitively identified by clinical criteria (Linhares *et al.*, 2001; Novikova *et al.*, 2002). Among methods used in yeast infections diagnosis, the most important are cultures of these microorganisms (Linhares *et al.*, 2001). The identification of *C. albicans* is based on a number of phenotypic characteristics, of which carbohydrate assimilation is of primary importance. For these purposes we used the commercial API 20 C AUX system, which is commonly used for diagnosis of *Candida* spp. (Ellabib *et al.*, 2001; Mercure *et al.*, 1996; Smith *et al.*, 1999; Wadlin *et al.*, 1999).

When microbial strain typing methods are compared, the most important characteristics are typeability, reproducibility and discriminatory power. The results of the present study confirm those of other authors, and namely that the API system shows good reproducibility, sensitivity and specificity (Wadlin *et al.*, 1999). However, the discriminatory power of API system is not high, and, if epidemiological investigation has to be performed, results of examination with the use of Api system should be supplemented with other methods. For example electrophoretic typing of esterases and biotyping based on proteolytic activity can be used as additional methods for differentiating strains. Other authors have already indicated the value of using a combined typing system (Quindos *et al.*, 1996).

We tentatively grouped the *C. albicans* strains into four categories, three with esterase patterns, and one without an esterase band. Because the *C. albicans* isolates show serological differences according to their geographical origin (Hannula *et al.*, 2001; Mercure *et al.*, 1996), serological examination of *C. albicans* seems to be very important from the epidemiological point of view. In our investigation, all examined strains belonged to the A serotype.

It must be pointed out that the application of esterase zymogram in bacterial taxonomy and epidemiology is often used (Branger *et al.*, 2003; Chetoui *et al.*, 1998; Gilot and Andre, 1995 and 1996; Giver *et al.*, 1998). The examination of properties of *C. albicans* enzymes is widely applied (De Bernardis *et al.*, 1990; Pichova *et al.*, 2001; Tsuboi *et al.*, 1996; Vazquez-Reyna *et al.*, 1999) and may be useful for epidemiological purposes. Tsuboi *et al.* 1996 showed that the induction level of *C. albicans* extracellular esterase was found to be correlated with fungal growth. Protease production is one of the most relevant factors of *Candida* spp. virulence in mucosal diseases, including vaginitis (De Bernardis *et al.*, 1990 and 1999; Kantarcioglu and Yucel, 2002; Pichova *et al.*, 2001; Rodrigues *et al.*, 1999; Smolenski *et al.*, 1997).

C. albicans is significantly more proteolytic than the non-albicans yeasts (Wu and Samaranyake, 1999). Vaginal candidiasis may be observed as an acute disease which is characterized by a presence of inflammation symptoms. The cause of development of vaginal candidiasis may be reinfection from a gastrointestinal reservoir, sexual transmission, or impaired host defence mechanisms and enhanced *Candida* virulence (Kubota, 1998). There is clear evidence that proteolytic *C. albicans* strains were more virulent than the non proteolytic ones (De Bernardis *et al.*, 1999). In our study, 95.7% of the investigated strains exhibited proteolysis, when albumin, albumin and casein or casein were used as substrates (Table I). Similar results of

proteolytic activity are reported by Kantarcioglu *et al.*, 2002, who showed that 95% of clinical *C. albicans* isolates exhibited proteolysis on media with bovine serum albumin. On the other hand, Odds *et al.* (1983) reported that only one-third of *C. albicans* strains isolated from the female genital tract produced proteinase against human albumin. In the present study, the analysis of albumin and casein proteolysis is used in biotyping of *C. albicans* isolates. We found that 23 (50%) of the strains exhibited lysis of both these proteins and only two strains did not reveal proteolytic activity. It seems probable, then that the examination of lipolytic activity are of minor importance in epidemiological investigation of vaginal *C. albicans*.

Literature

- al-Rawi N. and K. Kavanagh. 1999. Characterisation of yeasts implicated in vulvovaginal candidosis in Irish women. *Br. J. Biomed. Sci.* **56**: 99–104.
- Branger C., C. Gardye, J.O. Galdbart, C. Deschamps and N. Lambert. 2003. Genetic relationship between methicillin-sensitive and methicillin-resistant *Staphylococcus aureus* strains from France and from international sources: delineation of genomic groups. *J. Clin. Microbiol.* **41**: 2946–51.
- Branger C., P. Gouillet, A. Boutonnier and J.M. Fournier. 1990. Correlation between esterase electrophoretic types and capsular polysaccharide types 5 and 8 among methicillin-susceptible and methicillin-resistant strains of *Staphylococcus aureus*. *J. Clin. Microbiol.* **28**: 150–1.
- Chetoui H., E. Delhalle, P. Melin, M.J. Struelens, R. De Ryck, P. Osterrieth and P. De Mol. 1998. Typing of nosocomial strains of *Serratia marcescens*: Comparison of pulsed-field gel electrophoresis of macrorestriction fragments with biotyping, esterase typing and ribotyping. *Res. Microbiol.* **149**: 137–143.
- De Bernardis F., L. Agatensi, I.K. Ross, G.W. Emerson, R. Lorenzini, P.A. Sullivan and A. Cassone. 1990. Evidence for a role for secreted aspartate proteinase of *Candida albicans* in vulvovaginal candidiasis. *J. Infect. Dis.* **161**: 1276–83.
- De Bernardis F., F. Mondello, G. Scaravelli, A. Pachi, A. Girolamo and L. Agatensi. 1999. High aspartyl proteinase production and vaginitis in human immunodeficiency virus-infected women. *J. Clin. Microbiol.* **3**: 1376–80.
- Ellabib M.S. and I.A. El Jariny. 2001. *In vitro* activity of 6 antifungal agents on candida species isolated as causative agents from vaginal and other clinical specimens. *Saudi Med. J.* **22**: 860–3.
- Farina C., V. Malighetti, J.P. Lombart, M. Toyb, Z. Youssouf, S. Caligaris, A. Matteelli, G. Chatel, M. Arosio and G. Carosi. 2000. Yeasts from vaginal exudates in the Comoros Islands. *J. Mycol. Med.* **10**: 91–93.
- Gilot P. and P. Andre. 1995. Characterization of five esterases from *Listeria monocytogenes* and use of their electrophoretic polymorphism for strain typing. *Appl. Environmental Microbiol.* **61**: 1661–65.
- Gilot P. and P. Andre. 1996. Serotyping and esterase typing for analysis of *Listeria monocytogenes* populations recovered from foodstuffs and from human patients with listeriosis in Belgium. *J. Clin. Microbiol.* **34**: 1007–10.
- Giver L., A. Gershenson, P.O. Freskgard and F.H. Arnold. 1998. Directed evolution of a thermostable esterase. *Proc. Natl. Acad. Sci.* **95**: 12809–13.
- Gouillet P. and B. Picard. 1991. *Pseudomonas aeruginosa* isolate typing by esterase electrophoresis. *FEMS Microbiol. Lett.* **78**: 195–200.
- Goswami R., V. Dadhwal, S. Tejaswi, K. Datta, A. Paul, R.N. Haricharan, U. Banerjee and N.P. Kochupillai. 2000. Species-specific prevalence of vaginal candidiasis among patients with diabetes mellitus and its relation to their glycaemic status. *J. Infect.* **41**: 162–6.
- Haberland-Carrodeguas C., C.M. Allen, F.M. Beck, W.J. Buesching, S.L. Koletar and P. Sundstrom. 2002. Prevalence of fluconazole-resistant strains of *Candida albicans* in otherwise healthy outpatients. *J. Oral Pathol. Med.* **31**: 99–105.
- Hannula J., B. Dogan, J. Slot, E. Okte and S. Asikainen. 2001. Subgingival strains of *Candida albicans* in relation to geographical origin and occurrence of periodontal pathogenic bacteria. *Oral Microbiol. Immunol.* **16**: 113–8.
- Kantarcioglu A.S. and A. Yucel. 2002. Phospholipase and protease activities in clinical *Candida* isolates with reference to the sources of strains. *Mycoses* **45**: 160–5.
- Kubota T. 1998. Chronic and recurrent vulvovaginal candidiasis. *Nippon Ishinkin Gakkai Zasshi.* **39**: 213–8.
- Kurnatowska A.J. 1998. Activity of hydrolytic enzymes of *Candida albicans* strains isolated from patients with periodontal and membrane mucosae of oral cavity diseases. *Mycopathologia* **141**: 105–9.
- Linhares L.M., S.S. Witkin, S.D. Miranda, A.M. Fonseca, J.A. Pinotti and W.J. Ledger. 2001. Differentiation between women with vulvovaginal symptoms who are positive or negative for *Candida* species by culture. *Infect. Dis. Obstet. Gynecol.* **9**: 221–5.
- Magliani W., S. Conti, A. Cassone, F. De-Bernardis and L. Polonelli. 2002. New immunotherapeutic strategies to control vaginal candidiasis. *Trends Mol. Med.* **8**: 121–6.
- Marai W. 2001. Lower genital tract infections among pregnant women: a review. *East Afr. Med. J.* **78**: 581–5.
- Mending W., M. Pinto De Andrade, J. Gutschmidt, R. Gantenberg, W. Presber and G. Schonian. 2000. Strain specificity of yeasts isolated from different locations of women suffering from vaginal candidosis, and their partners. *Mycoses* **43**: 387–92.
- Mercure S., S. Senechal, P. Auger, G. Lemay and S. Montplaisir. 1996. *Candida albicans* serotype analysis by flow cytometry. *J. Clin. Microbiol.* **34**: 2106–12.

- Moraes P.S., S. de Lima Goiaba and E.A. Taketomi. 2000. *Candida albicans* allergen immunotherapy in recurrent vaginal candidiasis. *J. Investig. Allergol. Clin. Immunol.* **110**: 305–9.
- Novikova N., E. Yassievich and P.A. Mardh. 2002. Microscopy of stained smears of vaginal secretion in the diagnosis of recurrent vulvovaginal candidosis. *Int. J. STD AIDS.* **13**: 318–22.
- Odds F.C., A.B. Abbott, R.L. Stiller, H.J. Scholer, A. Polak and D.A. Stevens. 1983. Analysis of *Candida albicans* phenotypes from different geographical and anatomical sources. *J. Clin. Microbiol.* **18**: 849–57.
- Pichova I., L. Pavlickova, J. Dostal, E. Dolejsi, O. Hruskova-Heidingsfeldova, J. Weber, T. Ruml and M. Soucek. 2001. Secreted aspartic proteases of *Candida albicans*, *Candida tropicalis*, *Candida parapsilosis* and *Candida lusitanae*. Inhibition with peptidomimetic inhibitors. *Eur. J. Biochem.* **268**: 2669–77.
- Quindos G., V. Lipperheide, B. Barturen, R. Alonso, J. Bikandi, R. San-Millan, M. Tellaetxe, L. Ribacoba and J. Ponton. 1996. A new method of antibiotyping yeasts for subspecies discrimination and distribution in human clinical specimens. *Eur. J. Epidemiol.* **12**: 55–62.
- Rodrigues A.G., P.A. Mardh, C. Pina-Vaz, J. Martinez de Oliveira and A.F. da Fonseca. 1999. Is the lack of concurrence of bacterial vaginosis and vaginal candidosis explained by the presence of bacterial amines? *Am. J. Obstet. Gynecol.* **181**: 367–70.
- Smith M.B., D. Dunklee, H. Vu and G.L. Woods. 1999. Comparative performance of the RapID Yeast Plus System and the API 20C AUX Clinical Yeast System. *J. Clin. Microbiol.* **37**: 2697–8.
- Smolenski G., P.A. Sullivan, S.M. Cutfield and J.F. Cutfield. 1997. Analysis of secreted aspartic proteinases from *Candida albicans*: purification and characterization of individual Sap1, Sap2 and Sap3 isoenzymes. *Microbiology* **143**: 349–56.
- Staib F. 1965. Serum-proteins as nitrogen source for yeastlike fungi. *Sabouraudia* **4**: 187–193.
- Taylor B.N., C. Fichtenbaum, M. Saavedra, J. Slavinsky III, R. Swoboda, K. Wozniak, A. Arribas, W. Powderly and P.L. Fidel Jr. 2000. *In vivo* virulence of *Candida albicans* isolates causing mucosal infections in people infected with the human immunodeficiency virus. *J. Infect. Dis.* **182**: 955–9.
- Tsuboi R., H. Komatsuzaki and H. Ogawa. 1996. Induction of an extracellular esterase from *Candida albicans* and some of its properties. *Infect. Immun.* **64**: 2936–40.
- Uriel J. 1966. Method of electrophoresis in acrylamide-agarosa gels (in French). 1966. *Bull. Soc. Chim. Biol. (Paris)*. **48**: 969–82.
- Vazquez-Reyna A.B., P. Ponce-Noyola, C. Calvo-Mendez, E. Lopez-Romero and A. Flores-Carreón. 1999. Purification and biochemical characterization of two soluble alpha-mannosidases from *Candida albicans*. *Glycobiology* **9**: 533–7.
- Wadlin J.K., G. Hanko, R. Stewart, J. Pape and I. Nachamkin. 1999. Comparison of three commercial systems for identification of yeasts commonly isolated in the clinical microbiology laboratory. *J. Clin. Microbiol.* **37**: 1967–70.
- Werner H. 1966. Studies on the lipase activity in yeasts and yeast-like fungi (in German). *Zbl. Bakt. I Orig.* **200**: 113–124.
- Wu T. and L.P. Samaranyake. 1999. The expression of secreted aspartyl proteinases of *Candida* species in human whole saliva. *J. Med. Microbiol.* **48**: 711–20.