

Relationship Between Measles Outbreaks Based on Genetic Analysis of Measles Virus Genomes Detected in Patients in Poland Between 2006 and 2012

AGATA MAKÓWKA and BOGUMIŁA LITWIŃSKA

National Institute of Public Health-National Institute of Hygiene, Department of Virology, Warsaw, Poland

Submitted 5 May 2014; Revised 10 September 2014, Accepted 24 October 2014

Abstract

This study describes the molecular characterization of 56 MeV strains obtained from 56 patients in Poland from 2006 to 2012. The C-terminal fragment of nucleoprotein gene was analysed. It has been found out during 2006 and 2012 MeV strains circulating in Poland belonged to genotypes D4, D5, D6 and B3. The D4 strains isolated in Poland were different from any other D4 strain circulating at the same time in Europe, whereas all other MeV strains isolated during 2007–2012 were related to strains from other countries. The present data suggest that after 2006 the MeV strains were imported.

Key words: measles outbreaks in Poland, genetic analysis of MeV, phylogenetic relationship between MeV strains

The measles virus (MeV) belongs to the family Paramyxoviridae, sub-family Paramyxovirinae, genus Morbillivirus. The genome of MeV is a single stranded, negative-sense RNA. Measles is highly infectious disease, characterized by unspecific prodromal symptoms, Koplik's spots and maculopapular rash and fever (Delpuet *et al.*, 2012).

MeV is serologically monotypic, there is no difference in the reactivity of sera in the course of infection with various MeV. Therefore current vaccines against measles provide protection against all wild-type viruses. However, there are some differences in nucleotide sequences between the MeV strains (Kühne *et al.*, 2006). The differences within certain genes determine the diversity of MeV genotypes. On the basis of the sequence of C-terminal fragment of nucleoprotein gene MeV strains are divided into clades A-H and 21 genotypes (WHO, 2012a).

Phylogenetic analysis of wild measles virus strains (MeV) together with epidemiological information, allows identifying sources of infection and distinguishing measles outbreaks between caused by native strains and caused by imported strains from other countries (Shakya *et al.*, 2012). Implementation of such research helps to prove the progress of the measles elimination program.

In 2010 WHO adopted the goal to eliminate endemic measles in the European Region by 2015 (Steffens *et al.*,

2010). Poland is a member of the WHO and the measles elimination program has been realized for many years. An improvement of the epidemiological situation of measles in Poland has been observed in recent years, due to maintenance of high immunization coverage. Before the introduction in 1974 of immunization against measles in Poland the incidence rate was 300–400 per 100.000. After the introduction of vaccination the incidence of measles rapidly decreased to 0,18 per 100.000 in 2012 (Janaszek *et al.*, 2002). At present, vaccination coverage against measles is more than 95%.

Poland is approaching measles elimination. According to the definition, elimination is when endemic measles cases are absent in a given area for at least 36 months from the last known case. A number measles cases were reported in Poland during the last few years, but mainly due to measles importation by travelling people.

The objective of this study was genotyping and phylogenetic analysis of measles virus strains in Poland during 2006–2012, which allows obtaining information about the sources of wild strains of MeV.

The epidemiological information about measles cases during 2006–2012 in Poland (*e.g.* time of onset, vaccination status, past journeys) was collected by the Department of Epidemiology, National Institute of Public Health-National Institute of Hygiene, Warsaw.

In Poland 120 measles cases were reported in 2006, 37 in 2007, 97 in 2008, 115 cases in 2009, 13 in 2010, 38

* Corresponding author: A. Makówka, National Institute of Public Health-National Institute of Hygiene, Department of Virology, Warsaw, Poland; e-mail: amakowka@pzh.gov.pl

in 2011 and 70 in 2012. They were recorded taking into account the presence of specific IgM antibodies against MeV and epidemiological link to confirmed measles cases. Among the 56 examined patients, 25 (44,6%) MeV strains were derived from Roma people.

According to the epidemiological investigation, 1 measles case in 2006 was associated with the MeV strain imported from Russia, whereas measles cases in 2007 were associated with importation from Ukraine. Measles cases in the Roma community were caused by MeV strains imported from the UK (2008 and 2009) and Romania (2012). One case in 2012 was a Polish soldier, who returned from the military mission in Libya.

In 2006 measles cases were dispersed in various locations in Poland and there were only a few cases in individual foci, in contrast to other years, when measles cases formed distinct clusters.

A total of 343 clinical samples (blood-105, throat swabs-133, urine-104 and nose swab-1) were collected in Poland during 2006 and 2012 from measles suspected cases were examined. All these samples were obtained from cases reported to the WHO Measles Elimination Programme, according to WHO criteria (WHO, 2012b). The samples were sent to the Department of Virology at National Institute of Public Health-National Institute of Hygiene, Warsaw, which is the National Laboratory for Measles and Rubella in Poland within the Global Measles and Rubella Laboratory Network, WHO.

MeV RNA was extracted from the clinical samples using QIAamp Viral RNA Mini kit (Qiagen). For detection of viral RNA conservative fragment of nucleoprotein gene of 400 bp corresponding to positions 465–864 was amplified by the nested RT-PCR method (Tischer *et al.*, 2004). Measles virus RNA was detected in 159 samples collected from 78 patients and then 56 of them was used for sequencing and phylogenetic analysis.

The variable fragment of nucleoprotein gene was amplified and the sequencing was performed by using the internal primers in the second round of amplification. This method has been previously described by Santibanez *et al.* (Santibanez *et al.*, 2002). The nucleotide sequences of the viral cDNA were analyzed by Chromas (version 1.45). Sequences were aligned and analysed by CLC Sequence Viewer 6.5.3 software. Phylogenetic trees were constructed by means of the neighbour-joining method. Genotype assignment was carried by program NCBI Genotyping Tool available on the website NCBI, USA (Rozanov *et al.*, 2004). This program allows to compare the query sequence with the sequences of WHO MeV reference strains and other known MeV sequences, available in the PubMed database.

Among 78 confirmed by RT-PCR measles cases, only for 56 patients the genotyping and phylogenetic analysis of MeV strains were possible to complete.

Confirmation of the presence of MeV RNA in clinical specimens allowed carrying out the sequencing and phylogenetic analysis. The phylogenetic tree illustrates the relationship between MeV strains isolated in Poland and in other countries. It also includes WHO reference strains (Fig. 1).

Individual clades are formed by the MeV strains registered in Measles Nucleotide Surveillance, named according to WHO nomenclature and the MeV strains closely related to them that were not registered in any database. They were named by laboratory code number. The reference strains are named according to WHO nomenclature.

Genotyping revealed that MeV strains circulating in Poland between 2006 and 2012 belonged to genotypes D4 (49), D5 (1), D6 (5) and B3 (1). The presented results show that in 2006 the diversity of MeV genotypes was observed (D4 and D5) whereas in 2007 the genotype of MeV strains was homogenous (D6). MeV strains circulated in Poland during 2008 and 2012 were related to each other and belonged to genotype D4.

Endemic transmission of MeV is defined as a chain transmission that is continuous for more than 12 months in a given area. Imported MeV strains have virological and epidemiological links with strains from other countries.

Epidemiological data suggests that in 2006 measles cases were caused by an indigenous virus strain circulating in Poland, genotyped D4. These measles cases were not linked to any other D4 strains circulated at the same time. According to the studies carried out in collaboration with Robert Koch Institute in Berlin, MeV strains isolated in 2006 in Poland were different from other MeV D4 circulating at the same time in Europe and they were related to the MeV strains detected in 1998 in Australia (MVi/Vic.AUS/10.98 – information from WHO Measles/Rubella European Regional Reference Laboratory). In 2006 the genotype D4 was also observed in Romania, however the Polish strains (*e.g.* MVi/Warsaw.POL/28.06/) were not identical to the Romanian one (MVi/Bucharest.ROU/04.06/3) (Makówka, 2007). The results of these studies suggest that genotype D4 detected in Poland in 2006 was indigenous. This confirms the epidemiological data concerning the size dispersion and number of measles cases in particular foci in Poland in 2006. MeV strains belonging to genotype D4 were circulating during 2006 in many European countries (Kremer *et al.*, 2008).

In 2006 one person (MVs/Warsaw.POL/41.06) was infected with genotype D5 of MeV. Sequencing results confirmed that the strain of this case was identical with the strain detected in Russia (MVs/NizhnyNovogrod.RUS/16.07/) and was related to the strain circulating in Germany in 2007 (MVs/Hanover.DEU/21.07/). This

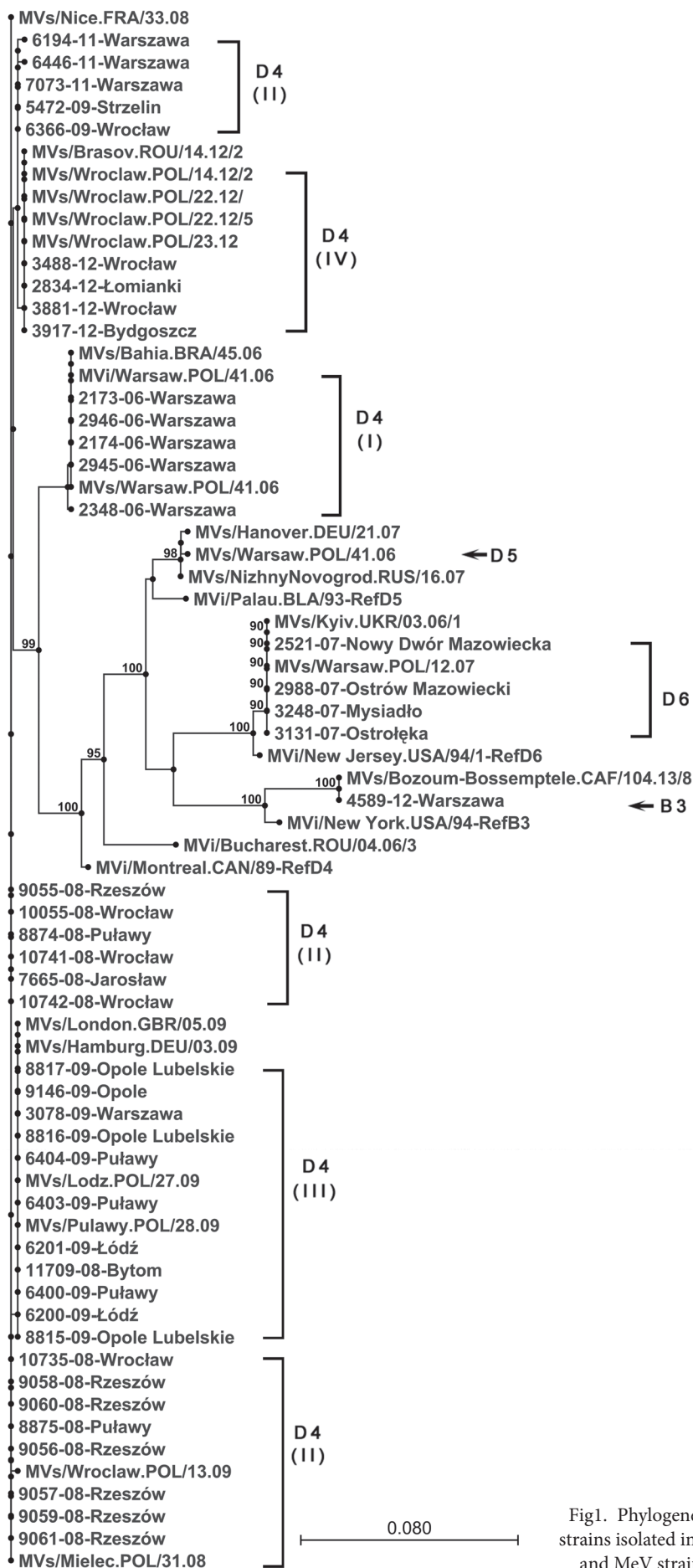


Fig1. Phylogenetic relationship between MeV strains isolated in Poland between 2006 and 2012 and MeV strains isolated in other countries.

confirmed the epidemiological investigation, which revealed that this sequence was isolated from a person who was diagnosed with measles just after returning from Russia.

In 2007 the genotype D4 in Poland was replaced by genotype D6. The phylogenetic analysis indicates the relationship with MeV strains D6 detected in 2005–2006 in Ukraine (MV/Kyiv.UKR/03.06/1), which was also confirmed by the epidemiological investigation. Therefore we conclude that measles in Poland in 2007 was imported from Ukraine (Spika *et al.*, 2006).

MeV strains isolated in 2008, 2009, 2011 and 2012 in Poland belonged to the genotype D4, however they differ from the strains detected in 2006. This suggests that there was a different source of MeV strains and a new transmission chain than in 2006. The phylogenetic analysis in this study indicates that MeV D4 strains, circulating in Poland in these years, were imported from other European countries: France (2008), Germany and Great Britain (2009) and Romania (2012).

The measles outbreaks in Poland in 2008, 2009 and 2012 were mainly observed in the Roma ethnic group. The MeV strain from 2008 is related to the strain from France (MV/Nice.FRA/33.08). The MeV strains circulated in 2009 were similar to the D4Hamburg strain (MV/Hamburg.DEU/03.09/) and to the strain from Great Britain (MV/London.GBR/05.09/). The MeV strain detected in 2012 was identical to the Romanian one (MV/Brasov.ROU/14.12/2).

The studies carried out with collaboration with the Robert Koch Institute in Berlin confirmed the results of the phylogenetic analyses. In clinical samples collected in 2009 from Polish patients the MeV strains identical to the D4-Hamburg strain were detected. There were strains isolated from the patients of Roma origin in Łódź, Puławy and Opole Lubelskie (Rogalska *et al.*, 2010; Mankertz *et al.*, 2011).

D4-Hamburg strain was imported from Great Britain to Germany at the end of the year 2008 and then to Bulgaria and caused more than 24 thousand measles cases (Marinowa *et al.*, 2009). It was the largest measles epidemic in Europe since the outbreak in Ukraine in 2006 (Velicko *et al.*, 2008). Furthermore, D4-Hamburg caused many measles cases lasting from 2008 to 2011 e.g. in Ireland, Austria, Greece, Serbia, Macedonia (Melidou *et al.*, 2011). Sporadic measles cases were also observed in Romania, Turkey, Switzerland and they were associated with Roma travellers from Bulgaria (Marinova *et al.*, 2009; Mankertz *et al.*, 2011; Muscat, 2011). Also in the Roma community in Romania some measles cases had been observed in 2004. There were more than 9 thousand measles cases registered and the circulation of D4-Bucharest strain till 2007 was the result of the travelling of Roma people (Kremer *et al.*,

2008). The strains D4-Bucharest and D4-Hamburg differ from each other, indicating two separate chains of MeV infection in the Roma population.

The measles outbreak occurred in the Roma community in Poland in 2012 was initiated by a child who had acquired measles in Romania. MeV strains belonging to genotype D4 were detected in Poland and this variant was found in Romania a few weeks before the beginning of outbreak in Poland (Necula *et al.*, 2013).

In 2012 one sequence belonging to genotype B3 (case 4589-12-Warszawa) was found in a sample derived from a soldier diagnosed with measles just after returning from Lybia and it was identical to the strain isolated in Central African Republic (MV/Bozoum-Bossemptele.CAF/104.13/8) (Fig. 1). Genotype B3 is frequently associated with measles imported from Africa to many European countries (Riddell *et al.*, 2005).

The results of genetic analysis shows that measles in Poland in 2006 was indigenous, whereas since 2007 measles in Poland was a result of the importation of MeV strains from other European countries. Between 2006 and 2012 there were 4 various MeV genotypes identified in Poland. In areas, where the indigenous MeV strains circulation is interrupted, the occurrence of diverse genotypes is observed, indicating different sources of importation of measles. In contrast, in endemic areas a limited number of MeV genotypes are detected.

The present results show that genotyping and phylogenetic analysis of MeV is an useful tool for outbreak investigation and Poland is on the right way to measles elimination. It is essential to continue the molecular surveillance of measles in Poland to monitor the pattern transmission of MeV.

Acknowledgements

This work was partially supported by Ministry of Science and Higher Education, Poland (NN 404 11 39 39).

Literature

- Delpout S., R.S. Noyce, R.W. Siu and C.D. Richardson.** 2012. Host factors and measles virus replication. *Current Opinion in Virology* 2: 773–783.
- Janaszek W., W. Gut and N.J. Gay.** 2002. Measles vaccine efficacy during an epidemic in 1998 in the highly vaccinated population of Poland. *Vaccine* 3446: 1–6.
- Kühne M., D.W. Brown and L. Jin.** 2006. Genetic variability of measles virus in acute and persistent infections. *Infect. Genet. Evol.* Jul 6(4): 269–276. Epub 2005 Sep 19.
- Kremer J.R., K.E. Brown, L. Jin, S. Santibanez, S.V. Shulga, Y. Aboudy, I.V. Demchyshyna, S. Djemileva, J.E. Echevarria, D.F. Featherstone and others.** 2008. High genetic diversity of measles virus, World Health Organization European Region, 2005–2006. *Emerg. Infect. Dis.* 14(1): 107–114.

- Makówka A., W. Gut and B. Litwińska.** 2007. Measles elimination programme on the world and Poland. (in Polish) *Przegl. Epidemiol.* 61(1): 135–142.
- Mankertz A., Z. Mihneva, H. Gold, S. Baumgarte, A. Baillet, R. Helble, H. Roggendorf, G. Bosevska, J. Nedeljkovic, A. Makowka and others.** 2011. Spread of measles virus D4-Hamburg, Europe, 2008–2011. *Emerg. Infect. Dis.* 17(8): 1396–401.
- Marinova L., M. Muscat, Z. Mihneva and M. Kojouharova.** 2009. An update on an ongoing measles outbreak in Bulgaria, April–November 2009. *Euro. Surveill.* Dec 17; 14(50).pii: 19442.
- Melidou A., G. Gioula, V. Pogka, M. Exindari, A. Moutoussi, D. Sgouras, K. Papadakis, D. Chatzidimitriou, D. Karabaxoglou, A. Mentis and others.** 2011. Molecular and phylogenetic analysis of Greek measles 2010 strains. *Epidemiol. Infect.* 31: 1–7.
- Muscat M.** 2011. Who gets measles in Europe? *J. Infect. Dis.* 204 Suppl 1: S353–365.
- Necula G., M. Lazar, A. Stanescu, A. Pistol, S. Santibanez, A. Mankertz and E. Lupulescu.** 2013. Transmission and molecular characterisation of wild measles virus in Romania, 2008 to 2012. *Euro. Surveill.* 12; 18(50): 20658.
- Riddell M.A., J.S. Rota and P.A. Rota.** 2005. Review of the temporal and geographical distribution of measles virus genotypes in the prevaccine and postvaccine eras. *Viol. J.* 22, 2: 87.
- Rogalska J., S. Santibanez, A. Mankertz, A. Makówka, L. Szenborn and P. Stefanoff.** 2010. Spotlight on measles 2010: An epidemiological overview of measles outbreaks in Poland in relation to the measles elimination goal. *Euro Surveill.* Apr 29; 15(17) pii.i: 19549.
- Rožanov M., U. Plikat, C. Chappey, A. Kochergin and T. Tatusova.** 2004. A web-based genotyping resource for viral sequences. *Nucleic Acids Res.* 1; 32 (Web Server issue): W654–9.
- Santibanez S., A. Tischer, A. Heider, A. Siedler and H. Hengel.** 2002. Rapid replacement of endemic measles virus genotypes. *J. Gen. Virol.* 83(Pt 11): 2699–2708.
- Shakya A.K., V. Shukla, H.S. Maan and T.N. Dhole.** 2012. Identification of different lineages of measles virus strains circulating in Uttar Pradesh, North India. *Viol. J.* 9: 237.
- Spika J.S., C. Aidyralieva, L. Mukharskaya, N.N. Kostyuchenko, M. Mulders, G. Lipskaya and N. Emiroglu.** 2006. Measles outbreak in the Ukraine, 2005–2006. *Euro. Surveill.* 11(3): E060309.1.
- Steffens I., R. Martin and P.L. Lopalco.** 2010. Spotlight on measles 2010: Measles elimination in Europe – a new commitment to meet the goal by 2015. *Euro. Surveill.* Volume 15, Issue 50.
- Tischer A., S. Santibanez, A. Siedler, A. Heider and H. Hengel.** 2004. Laboratory investigations are indispensable to monitor the progress of measles elimination—results of the German Measles Sentinel 1999–2003. *J. Clin. Virol.* 3: 165–178.
- Velicko I., L.L. Müller, R. Pebody, B. Gergonne, C. Aidyralieva, N. Kostyuchenko and J.S. Spika.** 2008. Nationwide measles epidemic in Ukraine: the effect of low vaccine effectiveness. *Vaccine* 2008 Dec 9; 26(52): 6980–5. doi: 10.1016/j.vaccine.2008.09.012. Epub Sep 19.
- WHO.** 2012a. Measles virus nomenclature update: 2012. *Wkly Epidemiol. Rec.* Mar 2, 87(9): 73–78.
- WHO.** 2012b. Library Cataloguing-in-Publication Data. Global measles and rubella strategic plan: 2012–2020.

